

Micromycetes on Surface of Some Imported Ripened Fruits



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Abstract

Investigations on surface mycoflora of imported fruits was conducted. It was revealed that the marketed imported fruits were contaminated by propagules of various micromycetes. The obtained results allow the conclusion that ripened fruit being juicy and rich sugar can become a good substrate for mycotoxin producing micromycetes. Some microorganisms especially of micromycetes belonging to the *Penicillium* Link, *Aspergillus* Mich. *Alternaria*. and other genera, are commonly found on fruits surface during investigation which are known to produce secondary metabolites (mycotoxins) of various compositions that are toxic to plants, animals and humans. Their detection frequency on the investigated fruit was determined.

Keywords: Mycoflora, Micromycetes, Contamination, Imported Fruit, Products.

Introduction

In terms of providing food with healthy nutrition, free from contaminants and long survived, the variety of fruits are imported from various countries. Regulations on contamination of micromycetes and mycotoxins have been set and strictly enforced by most importing countries. Imported fruit are juicy, rich vitamins, carbohydrates, organic acids, calcium, phosphorous and iron. All these constituents make these fruits vulnerable to post harvest attack by a variety of micromycetes to which they are resistant during the period of development on the plants.

Review of Literature

Most of the fungi are opportunists that may invade the harvested fruit directly or more usually through wounds attained during harvesting, transportation and poor handling practices(1). A few researchers have monitored the fungal spectra of fruits surface and their relative role in causing spoilage under storage (2). Many of these commodities facing a high risk of mycotoxin contamination. As imported fruits are often kept in unsuitable conditions like high humidity and temperature which is more favorable for microorganism spread, spoilage and mycotoxins contamination(8,9,10) which usually causes considerable economic losses . However, no work was done on regarding the surface micromycetes of imported fruits and their efficacy in spoilage in Jammu province(Jammu and Kashmir).

Aim of the Study

In view of this, investigation was conducted to monitor the surface micromycetes of imported fruits and its efficacy to causes spoilage was also evaluated by artificial inoculation.

Materials and Methods

Isolation of surface mycoflora

For isolation of the surface mycoflora associated with imported fruits, sample were taken in a sterilized polythene bags and brought to the laboratory and micromycetes were isolated by surface washing technique. In this method, 25 g of fruit peel was taken in a Erlenmeyer flask(250 ml containing 50 ml sterilized distilled water.) and subjected to horizontal shaking for 30 min. on rotary shaker. The liquid was then centrifuged at 3000rpm for 15 minutes and the residue thus obtained was mixed in 10 ml sterilized distilled water and shaken vigorously to obtain a homogenous suspension. This suspension was poured in sterilized petriplate at the rate of 2 ml/plate. Czapeks's Dox Agar medium (CDA) modified by adding streptomycin sulphate (0.06g/l) and rose Bengal (0.20g/l) was used and five replicates were maintained for each sample. The plates were incubated at 28+ 2°C for 7 days and the resulting fungal colonies subcultured on potato dextrose agar medium and identified.

Pathogenicity Test

It was conducted to check the ability of the recovered surface mycomycetes and assess their relative importance in the spoilage of fruits. For these tests, fruits of similar size and approximate of same maturity were surface sterilized with 70% alcohol and then incubated with the fungus by pin prick method (3). Inoculums used in each case was 5days old culture. After inoculation, the fruits were placed in sterilized glass chamber maintained at 28±2°C and relative humidity. Extent of spoilage was evaluated after 10 days of incubation.

Results and Discussion

Results depicted in table 1. that 33 micromycetes were found to be associated with the surface of fruits imported from abroad and sold in Jammu Market during 2017-18 i.e apple from California, Pohland; grapes from south Africa; Kiwi from Italy; Mango from Pakistan ; Orange from Egypt and Pear from China .

From Figure 1 It is observed that maximum number of micromycetes were reported on apple (25 species) and Pear (25 species) followed by mango(21 species), orange(20 species), grapes(17 species), Kiwi (17 species). Members of genus *Aspergillus* and *Penicillium* were predominant followed by *Fusarium* species. Other genera isolated as components of the micromycetes included *Alternaria*, *Botrytis*, *Curvularia*, *Cladosporium*, *Chaetomium*, *Colletotrichum*, *Emmericella*, *Geotrichum*, *Lasioidiplodia*, *Mucor*, *Phoma*, *Rhizopus*, *Trichothecium*, *Trichurus* and *Paecilomyces*. These genera may have reached the fruit surface directly from the orchard during picking and packing operation or they may have been carried along the packing material (6) .

Data presented in table 2 shows that most of the genera like *Alternaria alternate* (Fr) keissel, *Aspergillus flavus* link ex. Fries, *A. ochraceus* Wilhelm, *A.fumigatus*, *A.niger* Van Tiegham, *Botrytis cinere* Pers ex Fries, *Fusarium moniliforme* J.shed, *Lasioidiplodia theobromae* (Pat) Griff and Mauble , *Penicillium expansum* link ex Grey, *P. chrysogenum* Thom and *Rhizous stolonifer* (Ehrenb) Lind. were frequently present on the surface of all the fruits. Because of its frequently present on the fruit surface, known fast growing and have enough potential to cause fruit loss (5) ,these may cause deterioration of these respective fruits in the open market and even under cold storage. According to few researchers, propagule of these fungi causes fruit loss after injuries occurs on fruit surface (5).These injuries provide the entry of such mycopropagules resulting in rot development. In addition, as the fruits continues to respire even after harvesting, the resulted heat accelerates respiration and aging, which in turn makes the fruit susceptible to attack by the surface micromycetes (7).

After inoculation by their respective frequent occurred fungal species, it was also observed that maximum causes fruit rot when incubated at 28±2°C for 10 days with 100% relative humidity (table 3). In addition, among these fungal species, *Alternaria alternate*, *Aspergillus flavus*, *A. niger*, *Botrytis*

cinere, *F. moniliform*, *Lasioidiplodia theobromae*, *Penicillium expansum*, *P. chrysogenum*, and *Rhizopus stolonifer* showed maximum extent of rot in respective fruits while *Aspergillus ochraceus* cause minimum rot. *Aspergillus fumigatus* have not shown any rot may be not posses such specific macerating enzymes necessary for pathogenesis. Similarly , there are some reports on rot causing pathogens from apple (5), grapes(13), Kiwi(12),Mango(13) , Orange(14,15) and Pear(5) .

If fruits are damaged by such frequently occurred micromycetes and have ability to produce toxic metabolite, their product can be responsible to cause a severe and chronic diseases in human and animals. Similary, it is also reported in fruit by Santos et al.,2016. `Therefore from this study it is observed that there is more chance of availability of mycotoxins in our children's food and also lead to huge loss in finance from our state.

Conclusion

In view of this above study, it ardent need to check on mycoflora, uses of contaminated imported fruit in making their product. The aim of the present work was to monitor the occurrence of kinds of micromycetes on the surface of imported market fruits and these have potential to cause spoilage and to produce mycotoxins.

Endnotes

1. Dennis,C(1975).the microflora of the soft fruit (In : Preece,T.F Dickinson C.F editor. Microflora of aerial plant surface. London academic Press, 219-430.
2. Kalafatglue,H and Karepinar, M,(1989). Investiagation of spoilage microflora in selected apple cultivars during storage. University Ziraat dergisi, 26,347-356.
3. Tomkins , R.G and Trout, S.A(1931) the use of ammonium salt for the prevention of green mould in citrus. Jour. Pomol.Hort.Sci.,257-264
4. Singh Y.P and Sumbali, G (1998) . Pre- harvest microbial population of india jujube fruits . (Ziziphus mauritiana Lamk) and their implication in post harvest pathogenesis. Mycopathologia 142: 77-80
5. Singh Y.P and Sumbali, G (2004). Performance of apple and pear fruits cultivars to post harvest fruit rot fungi. Jour. Mycol. Pl. Pathol 34(3), 914-916.
6. Singh Y.P and Sumbali, G (2008). Studies on the role of surface micromycetes in causing post harvest rot of apple cv. Red delicious. Proc . Nat. Acad.Sci.India Sect. B, 78, 85-89.
7. Dasgupta M.K, Mandal N.C (1989). In: Dasgupta M.K , Mandal NC editor. Post harvest pathology of perishable, New Delhi and Mumbai: IBH Publishing CO. Pvt LTd, 1-623.
8. Cole, R.J. and Cox, R.H (1981). Hand book of toxic fungal metabolites – London ,
9. Janchova, D ., Lukova, L., Lebeda,A.,Pech,E, and Kutrova, E.(2000) Dynamic of enzymes during pathogenesois of *Fusarium oxysporium* on Pea seedling- Czech Mycology 52.141-142.

10. Lugauskas, A., Stakeniene, J. (2002). Toxin producing micromycetes on fruit, berries and vegetables. *Ann. Agric. Environ. Med.* 9: 183-197.
11. Santos, G., Mallos, L.M. and Morett, C.L. (2016). Quality and occurrence of mycotoxins in tomato product in the Brazilian market. *Enzyme Engineering* 5; 156.
12. Pennycook, S.R. (1985). Fungal fruit rots of *Actinidia deliciosa* (kiwi fruit). *New Zealand Journal of Experimental Agriculture* 13(4) 289-299.
13. Rathod, G.M. (2010). Survey of post harvest fungal diseases of some fruits from Marathwada regions of Maharashtra. *Indian Jour. Ecobiotechnol.* 2/6 7-10.
14. Bamba, R. and Sumbali, G. (2005). Co-occurrence of aflatoxin B1 and Cyclopiazonic acid (*Citrus aurantifolia* Swingle) during post harvest pathogenesis by *Aspergillus flavus*. *Mycopathologia* 159, 407-411.
15. Rasool, A., Zaheer, I. and Iram, S. (2014). Isolation and characterization of postharvest fungal pathogen of citrus varieties from the domestic markets of Rawalpindi and Islamabad. *Internat. Jour. Sci. Engin. Res.* 5(10) 408-418.

Table 1: Micromycetes Detected on the Imported Fruits from Jammu

S.no	Name of fruit	Imported from country	Micromycetes Species
1	Apple	California, Poland	<i>Alternaria alternata</i> (Fr.) Keissl. <i>Aspergillus flavus</i> link es. Fries <i>A. ochraceus</i> Wilhelm <i>A. sydowii</i> Thom and Church <i>A. fumigatus</i> Fresenius <i>A. niger</i> Van Tiegham <i>A. niveus</i> Blochwitz <i>A. solani</i> Sorauer <i>A. terrius</i> Thom <i>Botrytis cinerea</i> Pers. ex Fries <i>Curvularia lunata</i> <i>Cladosporium oxysporum</i> Kunze ex Fries <i>Colletotrichum gloeosporioides</i> (Penz) Sacc. <i>Emericella nidulans</i> (Eidam) Vuill <i>F. moniliforme</i> J. Sheld <i>F. verticilloides</i> (Sacc.) Nirenberg <i>Fusarium sporotricoides</i> Sherb. <i>Lasiodiplodia theobromae</i> (Pat) Griff and Manble <i>P. chrysogenum</i> Thom <i>P. pupureogenum</i> Stoll <i>P. citrinum</i> Thom <i>Paecilomyces variotii</i> Bainier <i>Penicillium expansum</i> link ex. Grey <i>Rhizopus stolonifer</i> (Ehrenb) Lind <i>Trichothecium roseum</i> (Pers) link es. Fries
2	Grapes (black)	South Africa	<i>Alternaria alternata</i> (Fr.) Keissl. <i>Aspergillus flavus</i> link es. Fries <i>A. niger</i> Van Tiegham <i>A. terrius</i> Thom <i>A. ochraceus</i> Wilhelm <i>A. fumigatus</i> Fresenius <i>Botrytis cinerea</i> Pers. ex Fries <i>Curvularia lunata</i> <i>Cladosporium oxysporum</i> Kunze ex Fries <i>Emericella nidulans</i> (Eidam) Vuill <i>F. moniliforme</i> J. Sheld <i>Geotrichum candidum</i> <i>Lasiodiplodia theobromae</i> (Pat) Griff and Manble <i>Penicillium expansum</i> link ex. Grey <i>P. chrysogenum</i> Thom <i>P. pupureogenum</i> Stoll <i>Rhizopus stolonifer</i> (Ehrenb) Lind
3	Kiwi	Italy	<i>Alternaria alternata</i> (Fr.) Keissl. <i>Aspergillus flavus</i> link es. Fries <i>A. ochraceus</i> Wilhelm <i>A. fumigatus</i> Fresenius <i>A. niger</i> Van Tiegham <i>A. terrius</i> Thom

			<i>Botrytis cinera</i> Pers.ex Fries <i>F. moniliforme</i> J.Sheld <i>Geotrichum candidum</i> <i>Lasiodiplodia theobromae</i> (Pat) Griff and Manble <i>P. brevicompactum</i> Dierckx <i>P. chrysogenum</i> Thom <i>P.italicum</i> Wehmer <i>Penicillium expansum</i> link ex. Grey <i>Rhizopus stolonifer</i> (Ehrenb) Lind <i>Trichothecium roseum</i> (Pers)link esx Fries <i>Trichurus spiralis</i> Hasselbring
4	Mango	Pakistan	<i>Alternaria alternate</i> (Fr.) keissl <i>Aspergillus flavus</i> link es.Fries <i>A. ochraceus</i> Wilhelm <i>A.fumigatus</i> Fresenius <i>A.niger</i> Van Tiegham <i>A.terrius</i> Thom <i>Botrytis cinera</i> Pers.ex Fries <i>Cladosporium oxysporum</i> Kunze ex Fries <i>Colletotrichum gloeosporioides</i> (Penz) sacc <i>Curvularia lunata</i> <i>F. moniliforme</i> J.Sheld <i>F. verticilliodes</i> (Sacc.) niren-berg <i>Fusarium sporotricoides</i> Sherb <i>Geotrichum candidum</i> <i>Lasiodiplodia theobromae</i> (Pat) Griff and Manble <i>Mucor heimalis</i> Wehmer <i>P. chrysogenum</i> Thom <i>P. italicum</i> Wehmer <i>P.citrinum</i> Thom <i>Penicillium expansum</i> link ex. Grey <i>Rhizopus stolonifer</i> (Ehrenb) Lind
5	Orange	Egypt	<i>Alternaria alternate</i> (Fr.) keissl <i>Aspergillus flavus</i> link es.Fries <i>A. ochraceus</i> Wilhelm <i>A.fumigatus</i> Fresenius <i>A.niger</i> Van Tiegham <i>Botrytis cinera</i> Pers.ex Fries <i>Emericella nidulans</i> (Eidam) Vuill <i>Fusarium sporotricoides</i> Sherb <i>F. moniliforme</i> J.Sheld <i>F. verticilliodes</i> (Sacc.) niren-berg <i>Geotrichum candidum</i> <i>Lasiodiplodia theobromae</i> (Pat) Griff and Manble <i>Mucor heimalis</i> Wehmer <i>P. chrysogenum</i> Thom <i>Penicillium expansum</i> link ex. Grey <i>P.citrinum</i> Thom <i>P. italicum</i> Wehmer <i>P.digitatum</i> <i>Rhizopus stolonifer</i> (Ehrenb) Lind
6	Pear	China	<i>Alternaria alternate</i> (Fr.) keissl. <i>Aspergillus flavus</i> link es.Fries <i>A. ochraceus</i> Wilhelm <i>A. sydowii</i> Thom and Church <i>A.fumigatus</i> Fresenius <i>A.niger</i> Van Tiegham <i>A. niveus</i> Blochwitz <i>A.terrius</i> Thom <i>Botrytis cinera</i> Pers.ex Fries <i>Curvularia lunata</i> <i>Cladosporium oxysporum</i> Kunze ex Fries

			<i>Colletotrichum gloeosporioides</i> (Penz) sacc. <i>Chaetomium globosum</i> Berkand Cult. <i>Emericella nidulans</i> (Eidam) Vuill <i>F. moniliforme</i> J.Sheld <i>F. verticillodes</i> (Sacc.) niren-berg <i>Fusarium sporotricoides</i> Sherb. <i>Geotrichum candidum</i> <i>Lasiodiplodia theobromae</i> (Pat) Griff and Manble <i>Penicillium expansum</i> link ex. Grey <i>P. chrysogenum</i> Thom <i>P.citrinum</i> Thom <i>P. pupurogenum</i> Stol <i>Phoma</i> sp <i>Rhizopus stolonifer</i> (Ehrenb) Lind <i>Trichothecium roseum</i> (Pers)link esx Fries
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Table 2: Micromycetes frequently occurred in imported fruits

S.No.	Micromycetes	Apple	Grapes	Kiwi	Mango	Orange	Pear
1	<i>Alternaria alternate</i> (Fr.) keissl.	+	+	+	+	+	+
2	<i>A.solani</i> Sorauer	+	--	-	-	-	-
3	<i>Aspergillus flavus</i> link es.Fries	+	+	+	+	+	+
4	<i>A. ochraceus</i> Wilhelm	+	+	+	+	+	+
5	<i>A. sydowii</i> Thom and Church	+	-	-	-	-	+
6	<i>A.fumigatus</i> Fresenius	+	+	+	+	+	+
7	<i>A.niger</i> Van Tiegham	+	+	+	+	+	+
8	<i>A. niveus</i> Blochwitz	+	-	-	--	--	-
9	<i>A.terius</i> Thom	+	+	+	+	-	+
10	<i>Botrytis cinera</i> Pers.ex Fries	+	+	+	+	+	+
11	<i>Curvularia lunata</i>	+	+	-	+	-	+
12	<i>Cladosporium oxysporum</i> Kunze ex Fries	+	+	-	+	+	+
13	<i>Colletotrichum gloeosporioides</i> (Penz) sacc.	+	-	-	+	-	+
14	<i>Chaetomium globosum</i> Berkand Cult.	-	-	-	-	-	+
15	<i>Emericella nidulans</i> (Eidam) Vuill	+	+	-	-	+	+
16	<i>Fusarium. moniliforme</i> J.Sheld	+	+	+	+	+	+
17	<i>F. verticillodes</i> (Sacc.) niren-berg	+	-	-	+	+	+
18	<i>Fusarium sporotricoides</i> Sherb.	-	-	-	+	+	+
19	<i>Geotrichum candidum</i>	-	+	+	+	+	+
20	<i>Lasiodiplodia theobromae</i> (Pat) Griff and Manble	+	+	+	+	+	+
21	<i>Mucor hiemalis</i> Wehmer	-	-	-	+	+	-
22	<i>Penicillium expansum</i> link ex. Grey	+	+	+	+	+	+
23	<i>P. chrysogenum</i> Thom	+	+	+	+	+	+
24	<i>P. pupurogenum</i> Stoll	+	+	-	-	-	+
25	<i>P. brevicompactum</i> Dierckx	+	-	+	-	--	-
26	<i>P. citrinum</i> Thom	+	--	-	+	+	+
27	<i>P. digitatum</i>	-	-	-	-	+	-
28	<i>P. italicum</i> Wehmer	-	--	+	+	+	-
29	<i>Phoma</i> sp.	-	-	-	-	-	+
30	<i>Rhizopus stolonifer</i> (Ehrenb) Lind	+	+	+	+	+	+
31	<i>Trichothecium roseum</i> (Pers)link esx Fries	+	-	+	-	-	+
32	<i>Trichurus spiralis</i> Hasselbring	--	-	+	-	-	-
33	<i>Paecilomyces variotii</i> Bainier	+	-	--	-	-	-

■ Figure 1. Occurrence of micromycetes on surface of fruits

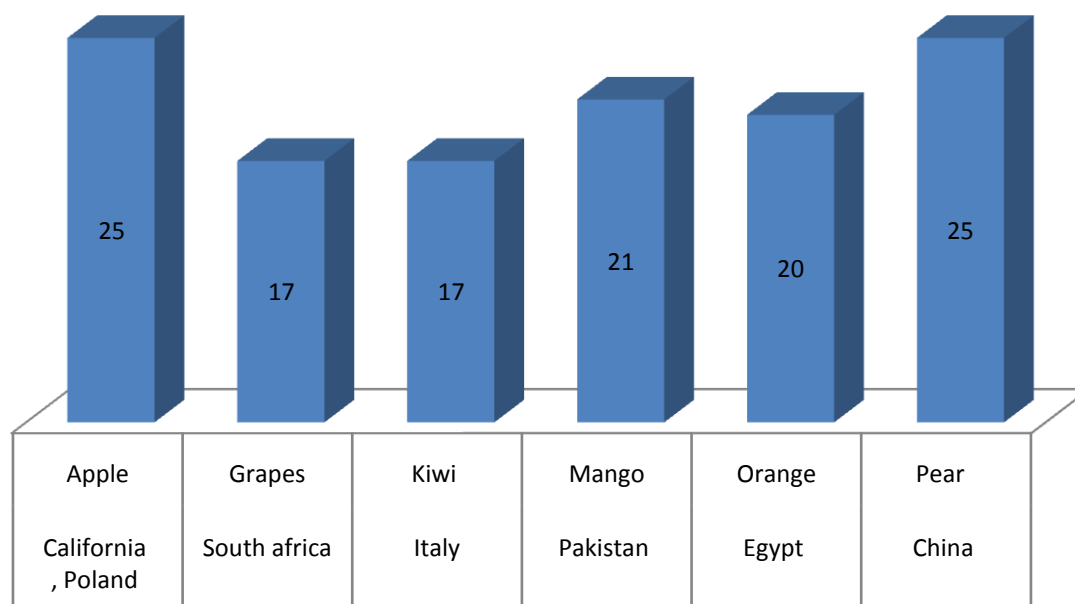


Table 3: Extent of Fruit Rots after Inoculation by Micromycetes

S. No	Name of species	Extent of fruit loss after incubation of 10 days at 28±2°C					
		Apple	Grapes	Kiwi	Mango	Orange	Pear
1	<i>Alternaria alternate</i> (Fr.) keissl.	+++	+++	++	+++	+++	+++
2	<i>Aspergillus flavus</i> link es.Fries	+++	+++	+++	+++	+++	+++
3	<i>A. ochraceus</i> Wilhelm	+	+	+	+	+	+
4	<i>A.fumigatus</i> Fresenius	-	-	-	-	-	-
5	<i>A.niger</i> Van Tiegham	+++	+++	+++	+++	+++	+++
6	<i>Botrytis cinera</i> Pers.ex Fries	+++	+++	+++	+++	+++	+++
7	<i>Fusarium. moniliforme</i> J.Sheld	+++	+++	+++	+++	+++	+++
8	<i>Lasiodiplodia theobromae</i> (Pat) Griff a9nd Manble	+++	+++	+++	+++	+++	+++
9	<i>Penicillium expansum</i> link ex. Grey	+++	+++	+++	+++	+++	+++
10	<i>P. chrysogenum</i> Thom	+++	+++	+++	+++	+++	+++
11	<i>Rhizopus stolonifer</i> (Ehrenb) Lind	+++	+++	+++	+++	+++	+++

+++ = maximum rot, + = minimum rot, - no rot